**Purpose**

To establish a procedure on how to triage and gross lymph nodes and spleens for lymphoma workup.

**Materials**

* 10% neutral buffered formalin
* RPMI media-located in refrigerator of frozen room 1 and/or central distribution
* Cytogenetics media located in refrigerator of frozen room 1 and/or central distribution
* Copies (x 2) of surgical pathology requisition
* Cytogenetics and flow requisitions, properly labeled and filled out-located above scale in frozen room 1
* Lymphoma tissue protocol sheet, properly labeled and filled out located above scale in frozen room 1.
* Requisition sticker (x 3) and specimen stickers (x2)
* Snap tube (x 1)-located on tissue procurement bench in frozen room 1

**Note:** Pre-made kits for Lymphoma workup will be available in the fridge in frozen room 1. Each kit will contain the following: lymphoma tissue protocol sheet (“yellow sheet”); prefilled container for flow cytometry with RPMI; prefilled cytogenetics media tube; cytogenetics requisition, plastic mold for snap freezing; 2 biohazard bags.

**Procedure**

Note: Lymphoma work-ups should be performed fresh and immediately if possible. If not, place in refrigerator for no more than 1 hour.

**Soft Tissue/Lymph Node Core Biopsy**

**Lymphoma Work up should not be done for core biopsies.**

1. All core specimens submitted to histology should be placed in formalin for routine formalin

processing.

**Lymph Node Biopsy on Whole or Partial Lymph Nodes**

1. Slice lymph node(s) into sections 2-3 mm thick perpendicular to the long axis.

2. If adipose or connective tissue is attached, set aside in specimen container and add formalin.

3. Submit at least one histologic section for routine formalin processing (cassette A1). If it is a large node, sample one slice per cm for histological analysis. If one part of the node grossly looks different (necrotic, etc), make sure you sample that part too. Note: These should be your cassettes A2 and so on.

4. Submit one piece of tissue for flow cytometry (at least 5 X 5 X 2 mm). Try to make sure it isn’t

adipose/connective tissue. Place tissue in a prefilled container with RPMI media and

attach specimen identification sticker. Fill out the flow cytometry requisition (see example below) and add the appropriate SU requisition sticker in the upper right hand corner. Identify and fill in the appropriate “Specimen Type” and check the “Leuks Leukemia/Lymphoma Immunohenotyping” box. Make a copy of the surgical pathology requisition and place both requisitions and RPMI tube in specimen biohazard bag. Write flow cytometry on biohazard bag and send to central distribution

5. If tissue is left over, submit a piece of tissue (at least 5 x 5 x 2mm) for cytogenetics analysis.

Place tissue in prefilled cytogenetics media tube and label with specimen identification sticker.

Note: Try to avoid any unnecessary manipulation since it will contaminate the specimen.

Make a copy of the surgical pathology requisition. Fill out a cytogenetics requisition (see example below and add the appropriate SU requisition sticker in the upper right hand corner. In the box labeled “Constitutional/Genetics Diagnosis/Indications for testing, write code: **CGNLN**. In the box labeled “Tests Requested for Malignancy”, check “chromosome analysis”. Place both requisitions and cytogenetics tube in biohazard bag. Write cytogenetics on biohazard bag and take to central distribution.

6. If additional tissue exists, snap freeze tissue in plastic mold containing OCT. Wrap in foil, label with specimen identification sticker and place in HistoChill in frozen room 1. This tissue will be transferred daily by technical staff to the -80 freezer in accessioning, and will be collected weekly by the hemepath group for long-term -80 storage.

7. Any extra tissue (after liberal sampling) can be saved in formalin with adipose/connective

tissue, as long as it has been adequately sampled. 8. If questions arise, call the hematopathology attending or fellow listed on the schedule.

8. A lymphoma tissue protocol sheet (see example below) must be filled out with the appropriate patient information (please attach a SU requisition sticker in the upper left hand corner), gross description, time in formalin and any ancillary studies performed. This sheet must be scanned into SoftpathDX along with the requisition. Staple the sheet to the requisition and package with specimen. Be sure to write time in formalin on specimen container.

**Spleen**

1. Measure and weigh specimen. Note any hilar lymph nodes and separate out.

2. Slice specimen as thinly as possible and examine carefully for lesions.

3. Take gross photographs of any lesions.

4. Contact the attending the on call hematopathologist or fellow and ask what tissue they want

submitted.

In most cases, the attending will want the hilar lymph nodes submitted (if present) for both

histological analysis and ancillary studies. See above "lymph node biopsy" steps 3 through 8 for

details.





